SHORT COMMUNICATION

TAPROBANICA, ISSN 1800–427X. Vol. 14, No. 02 (2025): pp. 323–324.

Published by Research Center for Climate Change & Faculty of Mathematics & Natural Sciences, Universitas Indonesia, Depok 16424, INDONESIA.

© distributed under Creative Commons CC-BY 4.0

http://www.taprobanica.org

https://doi.org/10.47605/tapro.v14i2.389



OPEN ACCESS

On the partial *TLR21* gene from the feathers of birds, a chicken, and a duck

The evolutionary arms race between pathogens and host organisms has shaped diverse immune detection systems across species. Among the most studied innate immune components are Toll-like receptors (TLRs), which bridge environmental signals with host immune responses (El-Zayat et al. 2019). Originally discovered in Drosophila melanogaster as a regulator of embryonic development, the Toll gene was later found to have antifungal immunity functions. This laid the foundation for the discovery of vertebrate TLRs, including those in birds. TLRs recognize structurally conserved molecules of microbes, known as pathogenassociated molecular patterns (PAMPs), encompassing lipids, lipoproteins, proteins, and nucleic acids (Rehman et al. 2021). Avian TLRs are structurally conserved with their mammalian counterparts but display unique features, such as the presence of TLR21 in birds, which functionally resembles mammalian TLR9 by detecting unmethylated CpG motifs (Lai et al. 2019). This study focuses on detecting and characterizing the partial cDNA of TLR21 from Indonesian endemic/endangered six avian species, such as the Java sparrow, using feather samples. By employing non-invasive methods, this study opens avenues for immunogenetic research without harming the animals, especially critical for conservation efforts.

Feather samples were collected non-invasively from six avian species: KUB-1 native chicken (*Gallus gallus domesticus*), Alabio duck (*Anas platyrhynchos Borneo*), zebra dove (*Geopelia striata*), spotted dove (*Streptopelia chinensis*), ring dove (*Streptopelia risoria*), and Java sparrow (*Lonchura oryzivora*). Ethical

clearance was obtained from the Ethics Commission of Institut Teknologi Bandung (02/ KEPHP-ITB/10-2019). Total RNA was extracted from the calamus of downy feathers using the RNeasy Mini Kit (Qiagen), followed by reverse transcription using AccuPower CycleScript RT Premix. PCR amplification targeted a 148 bp region of TLR21 using species-conserved primers. Amplicons were visualized on 2% agarose gel, purified, and sequenced by First (Singapore). Sequence editing alignment were conducted using Chromas, BioEdit, and Clustal Omega. Homology analysis was performed using BLAST, while protein translation was performed using the ExPASy tool. A phylogenetic tree was constructed using MEGA7 via the Neighbor-Joining method with 1,000 bootstrap replicates, incorporating zebrafish (Danio rerio) TLR21 as an outgroup.

Electrophoresis revealed a distinct 148bp band for five of the six avian species (excluding zebra dove, whose sequencing was inconclusive). Java sparrow's electropherogram showed two bands, but only the target 148 bp band was analyzed further. BLAST analysis confirmed ~99% homology of the partial TLR21 cDNA with reference sequences (NM 001030558.3 and NP 001025729.3), except for the Alabio duck, which showed ~96.7% homology. Translation of cDNA yielded 31 amino acid fragments aligning with the partial TLR21 protein of G. gallus. Phylogenetic analysis grouped KUB-1 chicken closely with Broiler chicken, while Alabio duck and the other doves formed separate branches. The inclusion of zebrafish TLR21 confirmed the ancestral linkage of this gene within nonmammalian vertebrates.

This study successfully demonstrates the expression of TLR21 in the feathers of several Indonesian avian species using non-invasive sampling. This approach is particularly valuable

for research on vulnerable or endangered birds like the Java sparrow, where preserving life is paramount. Despite its short sequence length, the 148 bp fragment proved informative for phylogenetic comparison, supported by previous studies (Asvapathanagul & Olson 2017). Short conserved sequences are often sufficient to infer evolutionary relationships when full-length sequences are unavailable or impractical to obtain. The detection of TLR21 in feathers also underscores the potential of feathers as a reliable source of immune-related genetic material. Feather calamus, particularly during growth phases, contains rich cellular material conducive to RNA extraction (Brown et al. 2022). The pulp within the feather follicle has been shown to mirror immune responses in lymphoid tissues, contributing valuable information for studies in avian immunology (Schat & Skinner 2022). Evolutionary analysis revealed close homology between domestic chicken breeds and partial divergence among ducks and doves. This finding previous observations supports paraphyletic nature of avian TLR21 evolution (Suryohastari et al. 2023). The lack mammalian orthologs further reinforces the uniqueness of TLR21 in birds, reptiles, and fish, with functional similarities but divergent evolutionary histories (Zhang et al. 2014). While the Zebra dove sequence could not be included due to low-quality reads, future studies may attempt higher coverage or alternative primer sets to address this gap. Furthermore, a more complete understanding of TLR21 function and evolution in birds would benefit from full-length sequencing, functional assays, and expression profiling across developmental stages and tissues.

Acknowledgments. This research was funded by the Research, Community Service & Innovation Program (P3MI-ITB) and a scholarship from the Ministry of Religious Affairs of Indonesia. We thank D.V.M. Sodirun (Balai Veteriner Subang) for research support, and M. Pangestu (Monash University), Yufang Liu (Henan Normal University), and Arif N.M. Ansori (Kumamoto University) for reviewing the manuscript.

Literature cited

- Asvapathanagul, P. & B.H. Olson (2017). Improving qPCR methodology for detection of foaming bacteria by analysis of broad-spectrum primers and a highly specific probe for quantification of *Nocardia* spp. in activated sludge. *Journal of Applied Microbiology*, 122(1): 97–105.
- Brown, C.L.J., T. Montina & G.D. Inglis (2022). Microbiome analysis of avian feathers using high-throughput sequencing: the potential of non-invasive sampling for microbial ecology. *Scientific Reports*, 12: 3561.
- El-Zayat, S.R., H. Sibaii & F.A. Mannaa (2019). Toll-like receptors activation, signaling, and targeting: an overview. *Bulletin of the National Research Centre*, 43(1): 187.
- Lai, C.Y., Y.T. Su & Y.C. Lin *et al.* (2019). TLR21 activation induces innate immune responses in avian macrophages. *Developmental & Comparative Immunology*, 92: 48–55.
- Rehman, Z.U., R.Z. Abbas & M.K. Iqbal *et al.* (2021). Toll-like receptors (TLRs) and their role in immune responses against avian diseases. *Microbial Pathogenesis*, 150: 104702.
- Schat, K.A. & M.A. Skinner (2022). *Avian Immunology*, 3rd Edition. Academic Press, London: 752pp.
- Suryohastari, A., A. Artarini & A.M. Pratama *et al.* (2023). Phylogenetic analysis of TLR21 in poultry: evidence of evolutionary divergence in domestic species. *Asian Pacific Journal of Tropical Biomedicine*, 13(5): 207–214.
- Zhang, H., S. Zhang & T. Wang *et al.* (2014). TLR21 is a potential ancestral TLR in vertebrates. *PLoS ONE*, 9(10): e107857.

Submitted: 25 Nov 2024, Accepted: 25 Jun 2025 Subject Editor: Yufang Liu

S.H. Sumarsono^{1*}, R.R.B. Suryohastari², E.A. Giri-Rachman¹, S.A.R. Nisa³

School of Life Sciences & Technology, Bandung
 Institute of Technology, Bandung, Indonesia
 Department of Biology, Faculty of Sciences &
 Technology, Syarif Hidayatullah State Islamic
 University Jakarta, Indonesia
 Faculty of Biology, Jenderal Soedirman University,
 North Purwokerto, Indonesia
 *E-mail: sonyheru@sith.itb.ac.id